

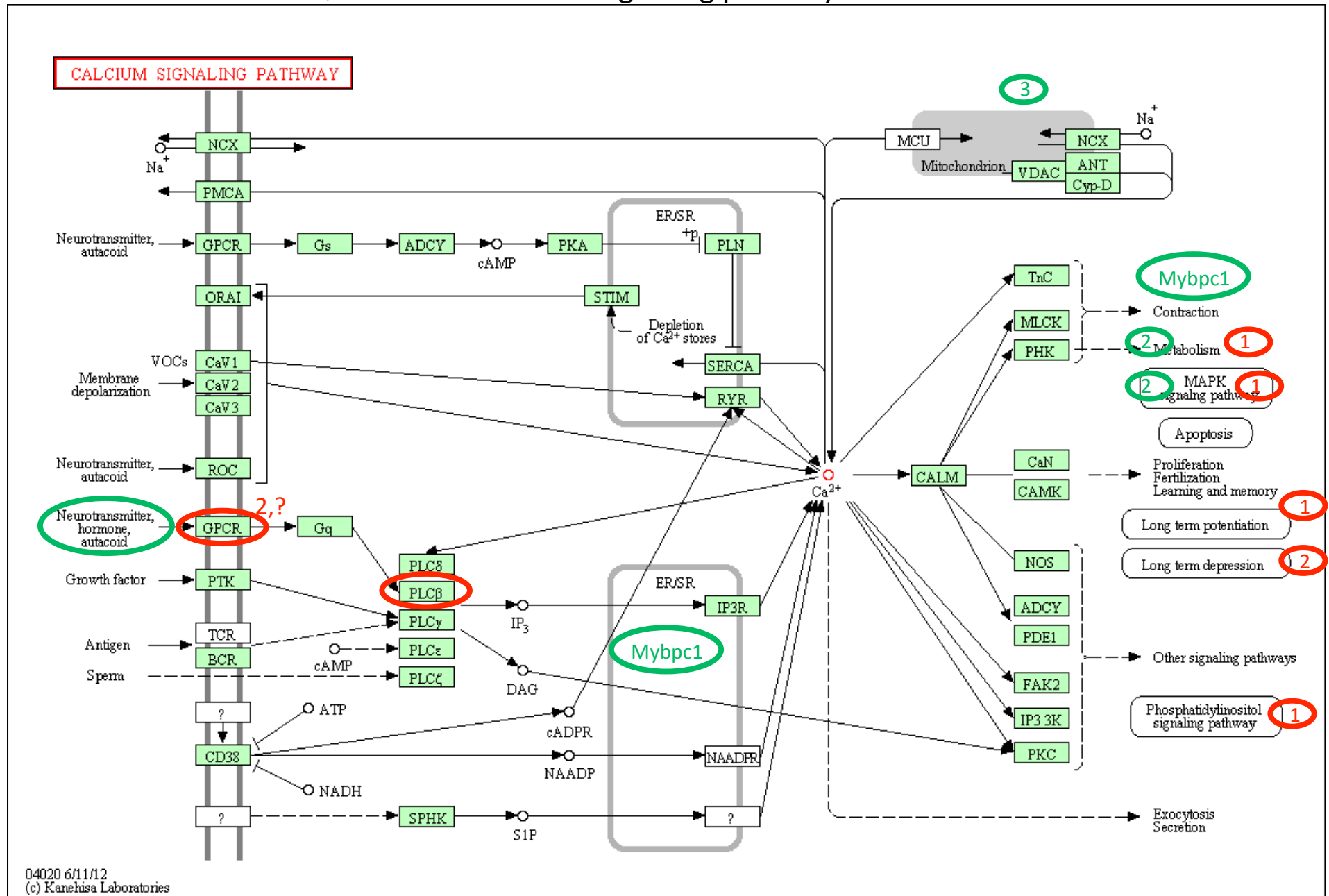
Lab meeting 8-17-12

- See spreadsheets

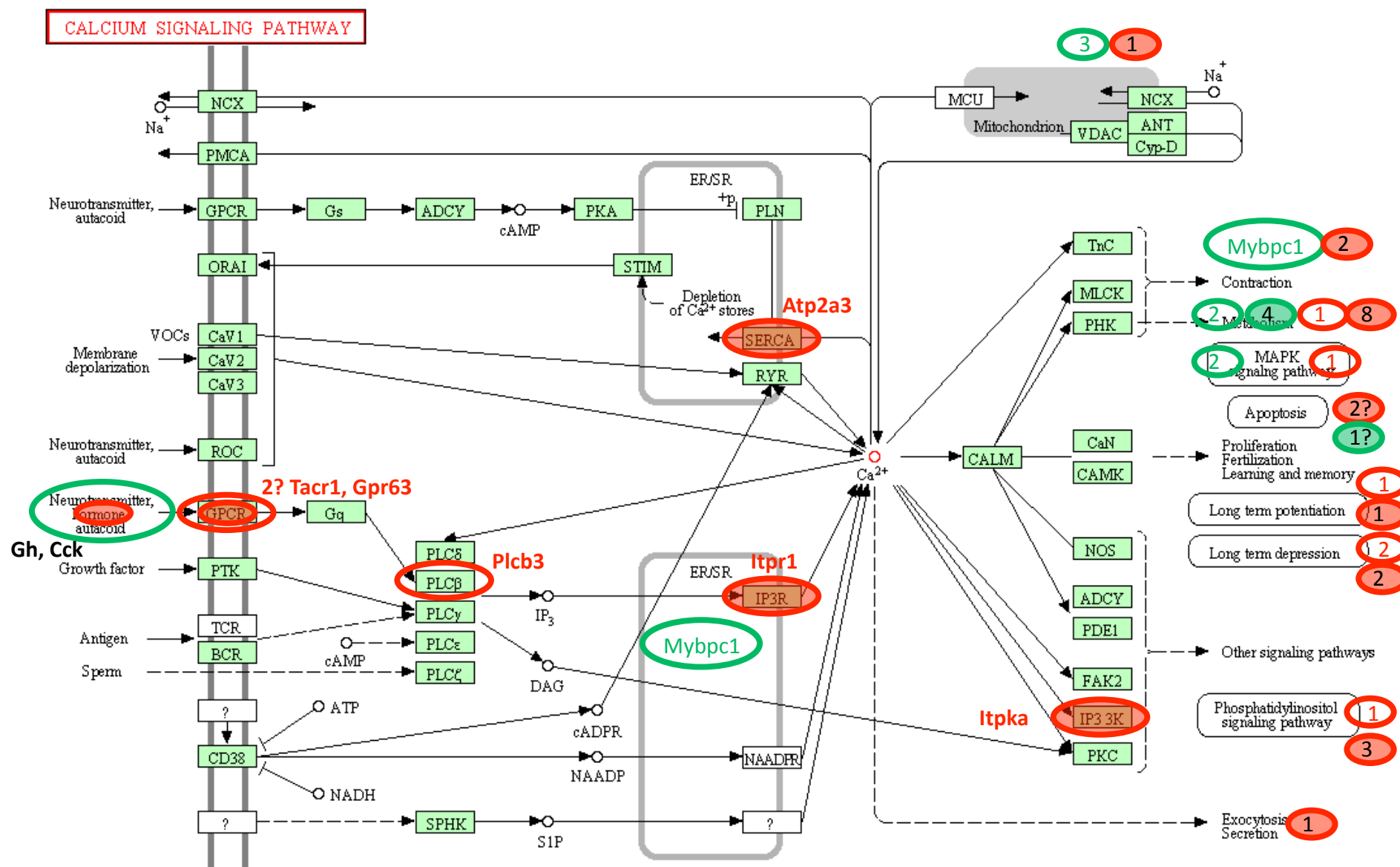
Comparison of RNA expression patterns in some well known pathways (Q127 vs WT)

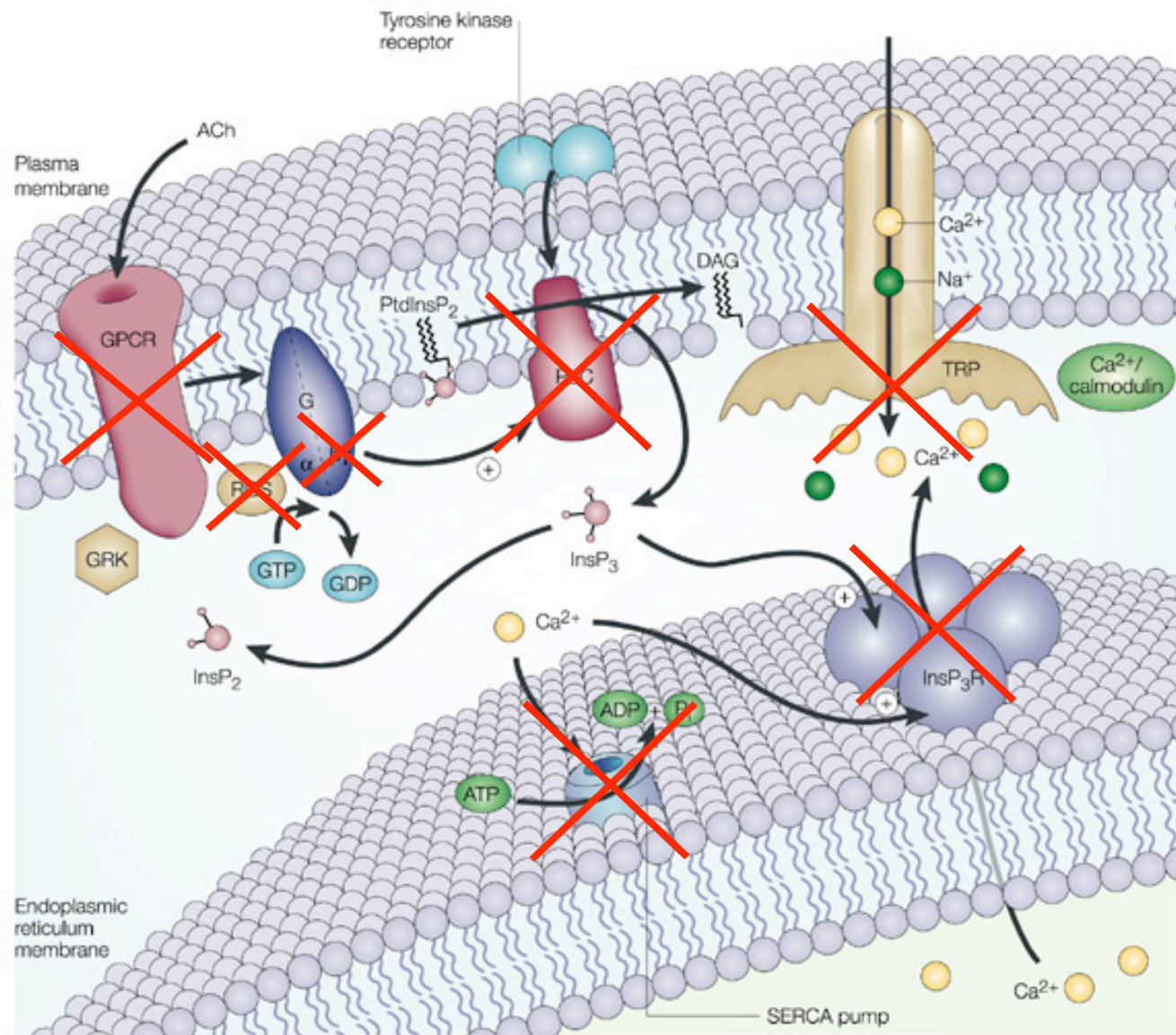
Key: A significant difference in expression is represented by a circle. Red signifies lower expression relative to WT, green signifies higher expression. Open circles represent 4 week-old comparisons, filled circles represent 8 week-old comparisons.

Q127 effects on Ca⁺⁺ signaling pathway at 4 weeks



Q127 effects on Ca⁺⁺ signaling pathway at 8 weeks

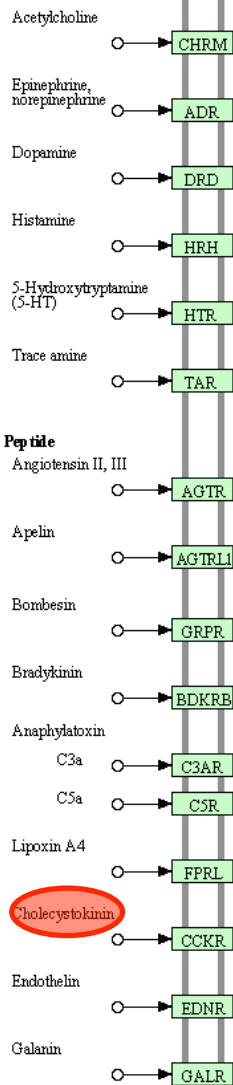




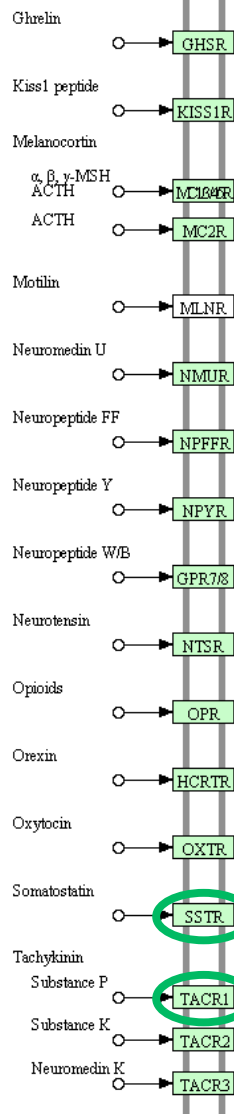
NEUROACTIVE LIGAND-RECEPTOR INTERACTION

GPCRs

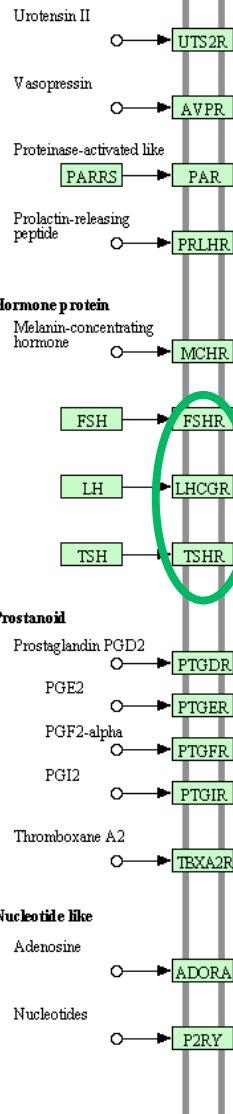
Class A Rhodopsin like Amine



Peptide



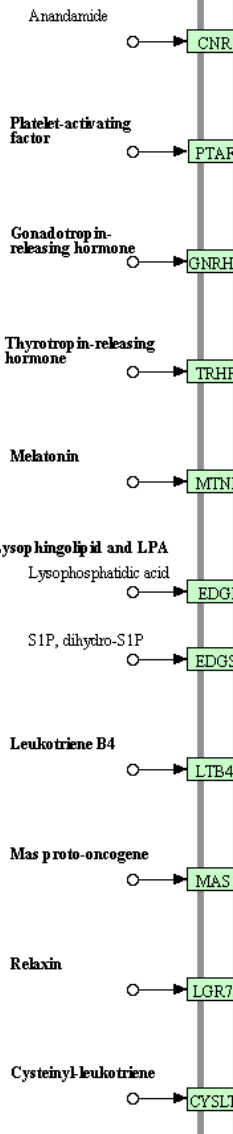
Hormone protein



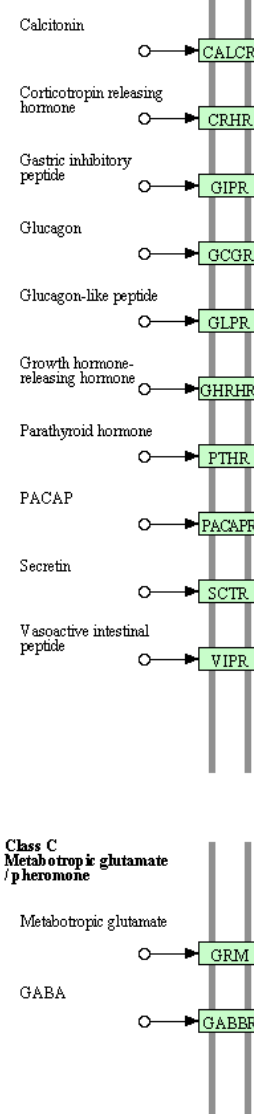
Prostanoid

Nucleotide like

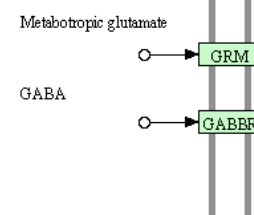
Cannabinoid



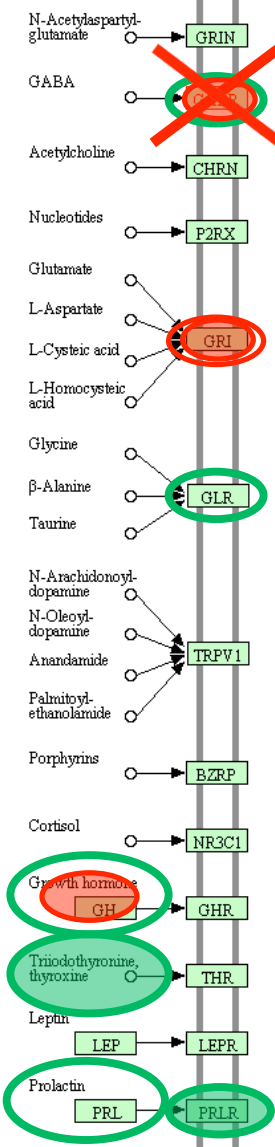
Class B Secretin like



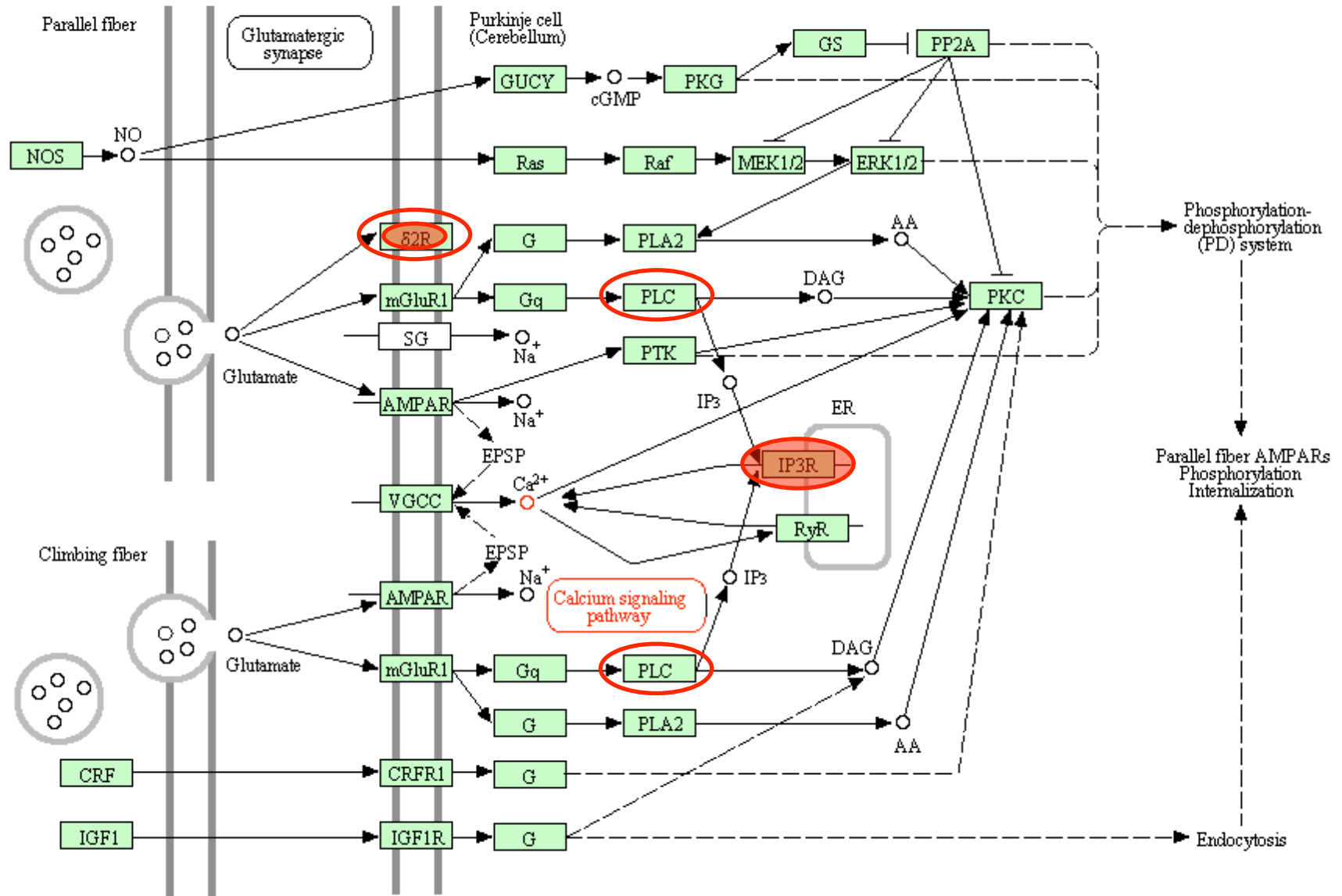
Class C Metabotropic glutamate / pheromone



Channels / other receptors



LONG-TERM DEPRESSION



GLUTAMATERGIC SYNAPSE

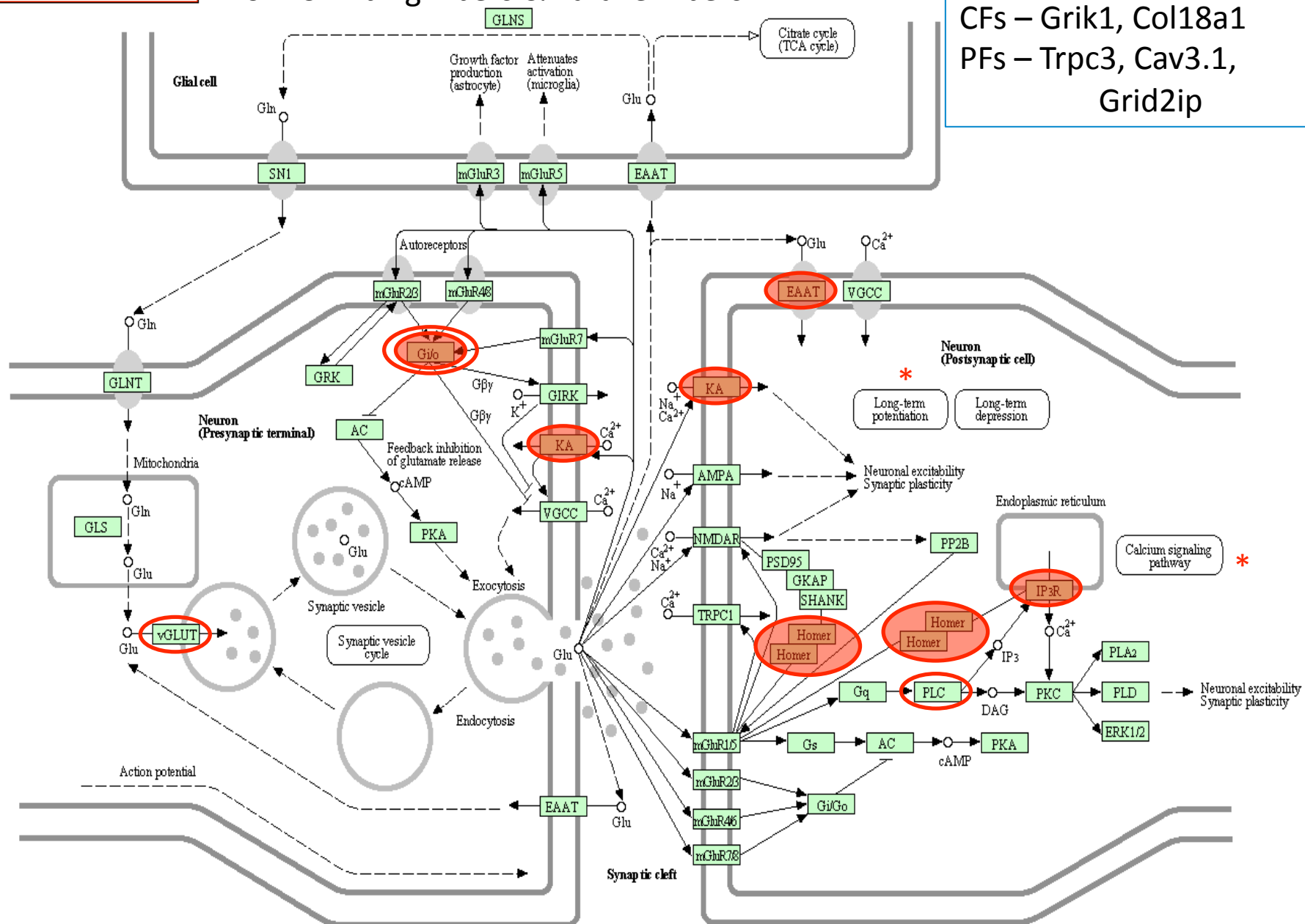
From Climbing Fibers & Parallel fibers

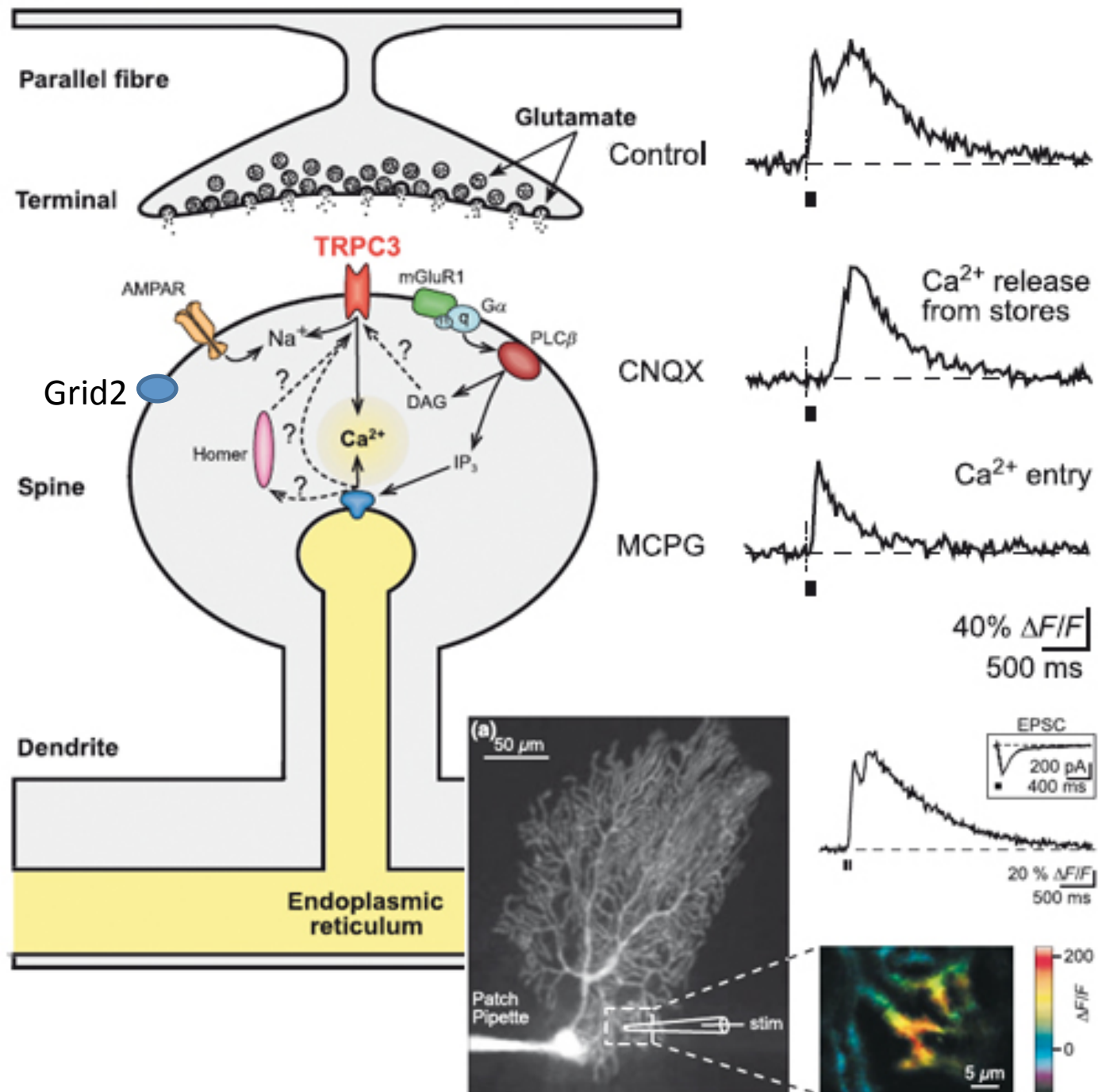
Genes down-regulated:

CFs – Grik1, Col18a1

PFs – Trpc3, Cav3.1,

Grid2ip

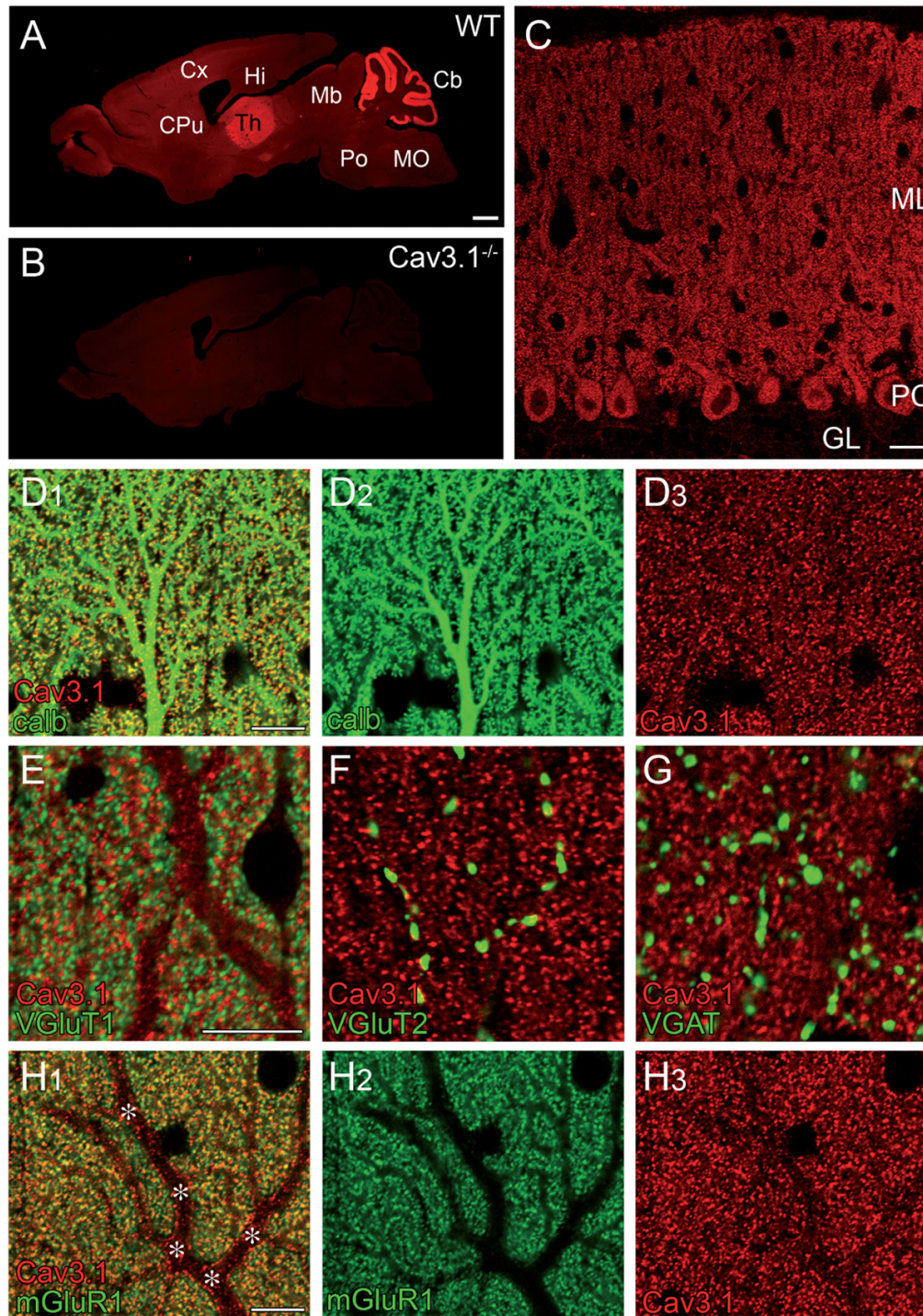




[Acta Physiol \(Oxf\)](#). 2008 Oct 28. [Epub ahead of print]

Mechanisms of metabotropic glutamate receptor-mediated synaptic signaling in cerebellar Purkinje cells.

[Hartmann J](#), [Konnerth A](#).

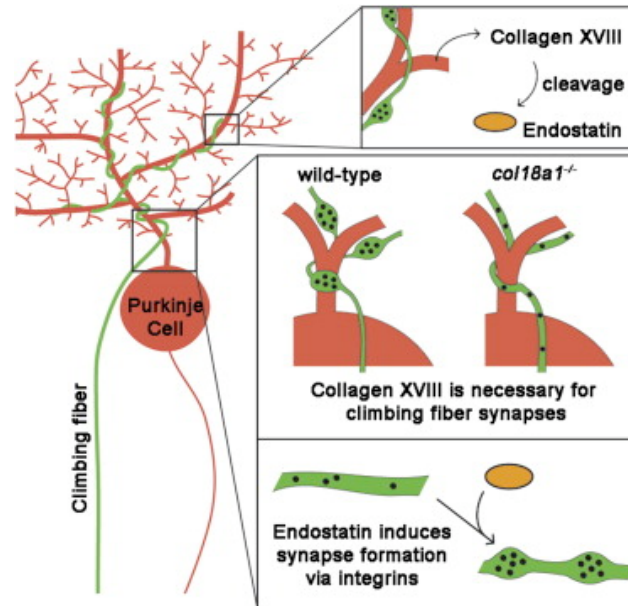


Cav3.1 slow type are thought to set pacemaker rhythm. Parallel fiber stimulation activates mGluR1-potentiated T-type Ca⁺⁺ transients

**Functional Coupling between mGluR1 and Cav3.1 T-Type
Calcium Channels Contributes to Parallel Fiber-Induced Fast
Calcium Signaling within Purkinje Cell Dendritic Spines**

**Michael E. Hildebrand,^{1*} Philippe Isope,^{2*} Taisuke Miyazaki,⁶ Toshitaka Nakaya,⁶ Esperanza Garcia,¹ Anne Feltz,²
Toni Schneider,⁵ Juergen Hescheler,⁵ Masanobu Kano,³ Kenji Sakimura,⁴ Masahiko Watanabe,⁶ Ste´phane Dieudonne´,²
and Terrance P. Snutch¹**

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Target-Derived Matricryptins Organize Cerebellar Synapse Formation through $\alpha 3 \beta 1$ Integrins

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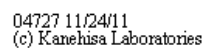
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From Basket cells



Research report

Identification of candidate Purkinje cell-specific markers by gene expression profiling in wild-type and *pcd*^{3J} mice

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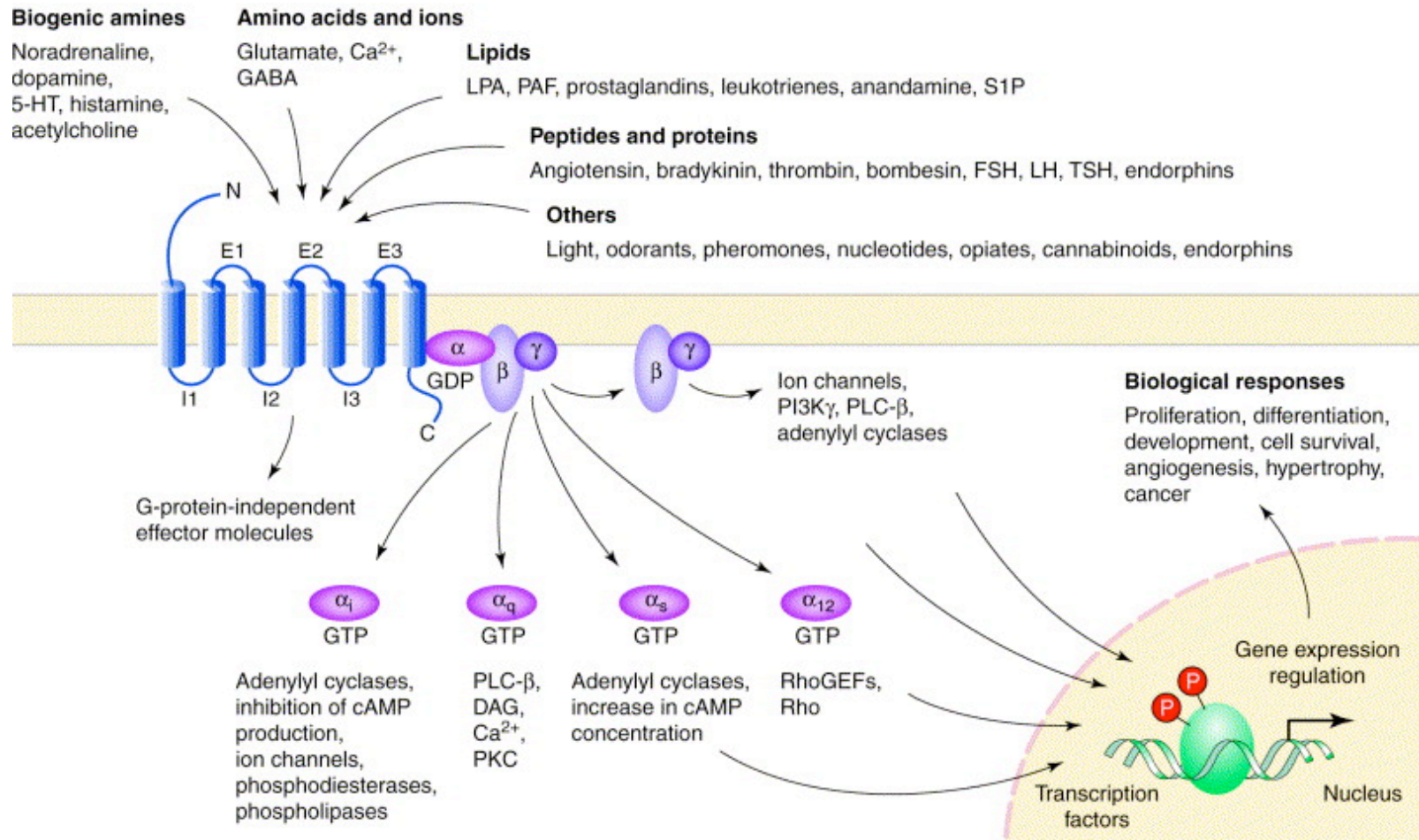
Accepted 7 October 2004

Abstract

The identification of mRNAs that have restricted expression patterns in the brain represents powerful tools with which to characterize and manipulate the nervous system. Here, we describe a strategy using microarray technology (Affymetrix Mouse Genome 430 2.0 Arrays) to identify mRNA transcripts that are candidate markers of cerebellar Purkinje neurons. Initially, gene expression profiles were compared between cerebella of 4-month-old Purkinje cell degeneration (*pcd*^{3J}) mice, in which most Purkinje cells had already degenerated and wild-type littermates with a normal complement of Purkinje neurons. Of 14,563 probe sets expressed in wild-type cerebellum, 797 showed a significant ($p < 0.0001$) reduction in *pcd*^{3J} mice. These probes could represent transcripts with varying levels of specificity for Purkinje cells as well as transcripts in other cell types that decline as a secondary consequence of Purkinje cell loss. Ranking of the probe signals revealed that well-known Purkinje cell-specific transcripts such as calbindin and L7/pcp2 clustered in a group that was <33% of wild-type levels. Therefore, to identify potentially new Purkinje cell-specific transcripts that cluster with the known markers, more stringent selection criteria were applied (<33% of wild-type signal and $p < 0.0001$). With these criteria, 55 independent transcripts were identified of which 33 were annotated genes and 22 were ESTs and RIKEN cDNAs. A literature search revealed that 25 of the 33 annotated genes were expressed in Purkinje cells, with no data being available on the other 8. Thus, the additional 8 annotated and 22 un-annotated genes are clustered with many genes expressed in Purkinje cells making them candidate markers. To confirm the microarray data, eight representative annotated genes were selected including five reported to be in Purkinje neurons and three for which no data was available. Semi-quantitative RT-PCR demonstrated reduced expression of all eight transcripts in cerebella from *pcd*^{3J} mice. The promoters of genes expressed selectively in subsets of neurons can be used to direct heterologous gene expression in transgenic mice and the more restricted the expression pattern the greater their utility. Therefore, microarray analysis was used to assess expression levels of all 55 transcripts in cerebral cortex, striatum, substantia nigra and ventral tegmental area. This permitted the identification of a set of genes whose promoters might have utility for selectively targeting gene expression to cerebellar Purkinje cells.

See common genes spreadsheet

- **Regulator of G-protein signaling 8** is a [protein](#) that in humans is encoded by the *RGS8* [gene](#).
[\[1\]\[2\]](#)
- This gene is a member of the regulator of G protein signaling (RGS) family and encodes a protein with a single RGS domain. Regulator of G protein signaling (RGS) proteins are regulatory and structural components of G protein-coupled receptor complexes. They accelerate transit through the cycle of GTP binding and hydrolysis to GDP, thereby terminating signal transduction, but paradoxically, also accelerate receptor-stimulated activation.[\[2\]](#)
- Regulator of G protein signaling (RGS) family members are regulatory molecules that act as GTPase activating proteins (GAPs) for G alpha subunits of heterotrimeric G proteins. RGS proteins are able to deactivate G protein subunits of the Gi alpha, Go alpha and Gq alpha subtypes. They drive G proteins into their inactive GDP-bound forms. Regulator of G protein signaling 4 belongs to this family. All RGS proteins share a conserved 120-amino acid sequence termed the RGS domain. Regulator of G protein signaling 4 protein is 37% identical to RGS1 and 97% identical to rat Rgs4. This protein negatively regulates signaling upstream or at the level of the heterotrimeric G protein and is localized in the cytoplasm.[\[1\]](#)



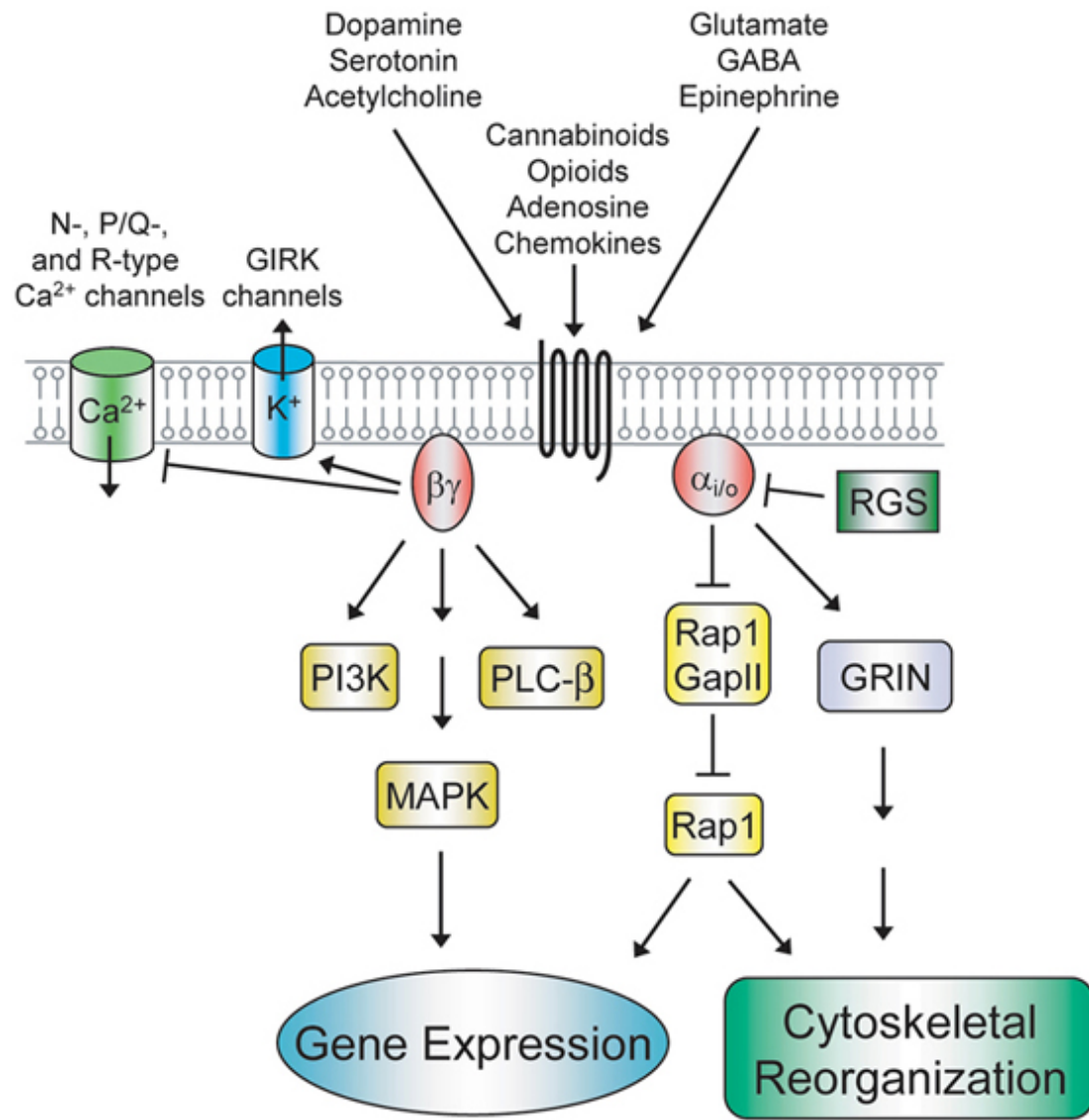


Figure 2. Effector pathways activated by $G_{i/o}$ signaling. Signals from a wide array of hormones, neurotransmitters, and chemokines are transduced into intracellular responses by $G_{i/o}$ -coupled receptors. Depicted are pathways that are stimulated by $G\alpha$ and $G\beta\gamma$ and lead to changes in gene expression and cytoskeletal reorganization. See text for further details. GIRK, G-protein-coupled inward rectifying potassium channels.