Original Investigation

Amyotrophic Lateral Sclerosis Risk for Spinocerebellar Ataxia Type 2 ATXN2 CAG Repeat Alleles A Meta-analysis

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IMPORTANCE Repeats of CAG in the ataxin 2 gene (*ATXN2*) in the long-normal range (sometimes referred to as *intermediate*) have been identified as modifiers of amyotrophic lateral sclerosis (ALS) risk. Prior studies have used thresholding considering various cutoffs for *ATXN2* repeat length.

OBJECTIVE To calculate association between *ATXN2* CAG repeat alleles and increased risk of ALS across multiple ethnic groups.

DATA SOURCES The MEDLINE database was searched for studies published by December 29, 2013, reporting *ATXN2* CAG repeat length in patients with ALS and controls.

STUDY SELECTION Studies were included if they reported original data on relative risks or odds ratios (ORs) from ALS and control populations for individual *ATXN2* alleles. Review articles that reported no new data were not included in the analysis.

DATA EXTRACTION AND SYNTHESIS Analysis of allele distribution was performed to ensure that all studies followed identical allele sizing. The ORs, 95% confidence intervals, and population attributable risk percentages were calculated according to standard procedures.

MAIN OUTCOMES AND MEASURES Occurrence of ALS associated with ATXN2 repeat alleles, expressed as ORs.

RESULTS Nine studies were analyzed, including 7505 controls and 6151 sporadic ALS cases. The ALS and control cohorts were recruited from different geographical and ethnic regions including the United States, French Canada/Canada, Belgium and the Netherlands, Germany, Italy, mainland China, Turkey, and Flanders-Belgium. The *ATXN2* CAG repeat lengths ranged from 13 to 39 in patients with ALS and from 13 to 34 in controls. The ORs were less than 1.00 for alleles with 25 to 28 repeats. The OR was 1.55 for 30 repeats, but this elevation was not statistically significant (95% CI, 0.88-2.73). The ORs were 2.70 (95% CI, 1.47-4.93) for 31 CAG repeats, 11.09 (95% CI, 4.16-29.57) for 32 repeats, and 5.76 (95% CI, 1.79-18.57) for 33 repeats.

CONCLUSIONS AND RELEVANCE In contrast to prior studies with smaller numbers, risk for ALS associated with long-normal alleles is complex. Alleles with 27 and 28 repeats lower ALS risk slightly. The risk for ALS increases beginning with 29 repeats and reaches a maximum at 32 and 33 repeats. Of note, alleles with repeats of these lengths are known to be predisposed to meiotic expansion to full-penetrance mutant alleles. In patients with ALS, alleles with 31 to 33 repeats may have undergone preferential expansion in motor neurons during mitosis or DNA repair. Our meta-analysis provides a framework for counseling individuals with long-normal *ATXN2* repeats.

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Corresponding Author: Stefan M. Pulst, MD, Department of Neurology, University of Utah, CNC Bldg, Fifth Floor, 175 N Medical Drive E, Salt Lake City, UT 84132 (stefan.pulst@hsc.utah.edu). S pinocerebellar ataxia type 2 (SCA2) is a rare autosomal dominant neurodegenerative disorder caused by expanded glutamine repeats located in the N-terminal region of the ataxin 2 gene (*ATXN2*; GenBank BC114546). The *ATXN2* polyglutamine allele length, although variable, is most commonly 22 repeats with rarer nonpathological alleles ranging from 14 to 31 repeats. Disease alleles causing ataxia contain 33 or more repeats.^{1,2} Longer pathogenic polyglutamine repeat lengths are inversely correlated with the age at onset.³ Pathology of SCA2 is characterized by considerable neuronal loss in the cerebellar Purkinje cell layer as well as all 4 deep cerebellar nuclei.⁴ In addition to cerebellar ataxia, patients with SCA2 may show other clinical signs such as saccade slowing, early hyporeflexia, severe tremors, and myoclonus.⁵

A recent study by Elden et al⁶ showed that *ATXN2* was a potent modifier of transactive response DNA-binding protein 43 kDa (TDP-43) through an RNA-dependent mechanism. TDP-43 has been found to be a major component of ubiquitinated cytoplasmic inclusions in neurons of patients with amyotrophic lateral sclerosis (ALS).^{7,8} Interestingly, Elden and colleagues went on to show that *ATXN2* alleles with 27 to 33 repeats, a class of alleles they designated as intermediate, were significantly associated with ALS. Since then, many studies⁹⁻¹⁶ have shown that long-normal *ATXN2* alleles contribute to an increased risk of ALS in a variety of ethnic backgrounds. Unfortunately, there seems to be a lack of consistency when defining the boundaries of intermediate-length repeats among the studies.

The purpose of our study was to perform a meta-analysis of published data to determine allele-specific risk for ALS for alleles ranging from 24 to 33 repeats.

Methods

Search Strategy

We undertook a MEDLINE database search on PubMed from July 2012 to December 2013 for studies that tracked *ATXN2* CAG repeat length in patients with ALS. The Medical Subject Heading terms used were SCA2 OR ataxin-2 OR ATXN2 AND ALS OR amyotrophic lateral sclerosis. The analysis was restricted to articles published in English. The final search was conducted on December 29, 2013. Studies were included if they reported original data, either directly in the publication or provided on request, on relative risks or odds ratios (ORs) from case and control populations, with cases defined as patients with ALS and control samples representative of the general population or controls who were matched for age and/or geographical region to the cases. We excluded review articles that reported no new data and studies that reported on interrupted *ATXN2* repeat lengths.

Statistical Analysis

Data were analyzed for genetic association between *ATXN2* and ALS using Mantel-Haenszel methods in Stata version IC/12.1 statistical software (StataCorp LP). The ORs were determined for allele counts greater than 23, taking into account the weight of each study. The ORs for alleles with more than 34 repeats could not be computed as these alleles were not found in con-

trols. A repeat of 0.5 was added to the control group so that ORs could be computed in instances in which controls were not reported for the respective repeat length. The addition of the small constant makes the estimate of Mantel-Haenszel pool ORs close to the maximum likelihood estimate.^{17,18} Population attributable risk percentages were calculated using standard methods.¹⁹

Study Populations

Most patients with ALS in previously published studies^{6,9-16} were diagnosed according to the El Escorial revised criteria.²⁰ In all examined studies, control samples were regionally matched to unrelated participants with no reported history of neurological disease.

Results

From the MEDLINE database search, PubMed yielded 28 articles, of which 14 fit the inclusion criteria for the metaanalysis. To be included within the meta-analysis, specific data for individual *ATXN2* allele frequencies greater than 23 for both the ALS and control cohorts were required. When sufficient data were found in the publication, the data were extracted from the published information. When adequate data were not published in the article, first and senior authors were contacted via e-mail to request additional information. Of the 14 eligible publications, 9 provided adequate data for analysis. Five of the 14 eligible studies did not respond to e-mails requesting additional data.

From the 9 studies, 7505 controls and 6151 sporadic ALS cases were analyzed. The ALS and control cohorts were recruited from different geographical and ethnic regions including the United States,⁶ French Canada/Canada,¹⁰ Belgium and the Netherlands,¹² Germany,¹¹ Italy,^{9,16} mainland China,¹⁵ Turkey,¹³ and Flanders-Belgium.¹⁴ The *ATXN2* CAG repeat lengths ranged from 13 to 39 in patients with ALS and from 13 to 34 in controls. Overall *ATXN2* CAG repeat length distribution is shown in **Figure 1** and in the eTable in the Supplement. Repeat lengths 23 or less represented 97.19% of alleles in the control population and 96.64% of alleles in patients with ALS. This is congruent with prior studies.^{1,21,22} As previously reported,^{22,23} the third most common repeat allele in controls and patients contained 27 repeats, suggesting that all studies used similar sizing standards, allowing comparisons across studies.

In contrast to prior studies that used a threshold for normal and predisposing alleles, which was often set post hoc, we had a sufficiently large number of alleles for each repeat size that allowed us to determine ORs specific for individual alleles. This meta-analysis revealed ORs of 1.00 or lower for alleles with 25 to 28 repeats. The first study examining ALS risk for CAG repeats between 27 and 33 reported a significantly increased risk for this class of alleles.⁶ The **Table** lists the ORs, 95% confidence intervals, and population attributable risk percentages for each allele summed across all studies. Contrary to previous studies, our analysis showed that ALS risk was decreased with CAG repeat lengths of 27 (OR = 0.77; 95% CI, 0.63-0.95) and 28 (OR = 0.44; 95% CI, 0.18-1.04). The OR was 1.55 for 30



Figure 1. Distribution of Ataxin 2 Gene (ATXN2) Polyglutamine Repeat Lengths in Both Amyotrophic Lateral Sclerosis and Control Cohorts

Table. Summary of Odds Ratios, 95% Confidence Intervals, and Population Attributable Risk Percentages for Ataxin 2 Gene Alleles With 24 to 33 CAG Repeats

CAG Repeats, No.	Odds Ratio (95% CI)	Population Attributable Risk %
24	1.10 (0.74-1.64)	0.0415
25	0.70 (0.39-1.25)	0.0113
26	0.80 (0.23-2.75)	-0.0003
27	0.77 (0.63-0.95)	-0.2434
28	0.44 (0.18-1.04)	-0.0306
29	1.15 (0.73-1.82)	0.0333
30	1.55 (0.88-2.73)	0.0616
31	2.70 (1.47-4.93)	0.1182
32	11.09 (4.16-29.57)	0.1948
33	5.76 (1.79-18.57)	0.0694

repeats, but this higher OR value did not indicate a statistically significant risk increase for ALS (95% CI, 0.88-2.73). Beginning with 31 repeats, ORs were significantly elevated (Table). The population attributable risk percentage for alleles with more than 30 repeats, however, was relatively low in keeping with the overall rare occurrence of these alleles in the general population (Table).

The ORs of each study for alleles of 30, 31, 32, and 33 repeats are shown in **Figure 2**. The ORs were 2.70 (95% CI, 1.47-4.93) for 31 CAG repeats, 11.09 (95% CI, 4.16-29.57) for 32 repeats, and 5.76 (95% CI, 1.79-18.57) for 33 repeats. Not all studies were informative for each allele class, when the respective allele was not present in cases and in controls. This is particularly true for alleles of more than 34 repeats that were found in patients with ALS but not in controls (Figure 1).

Discussion

Our analysis used 9 studies from the previously reported literature. Our meta-analysis supports previous observations of Distributions of *ATXN2* polyglutamine repeat lengths are shown for 23 or fewer CAG repeats (A) and for at least 24 CAG repeats (B).

an increased risk of ALS with certain expanded *ATXN2* alleles. This meta-analysis, however, provides a differentiated picture with some of the intermediate alleles associated with reduced risk and others with increased risk.

Elden et al⁶ originally defined the SCA2 repeat length for increased ALS risk to be 27 or more repeats. Since that 2010 publication, the clinically relevant range has been defined as widely as 29 or more repeats¹² or more restricted to 31 to 34 repeats.¹¹ Our meta-analysis has further defined the clinically relevant range of expanded *ATXN2* polyglutamine regions that increase ALS risk as 31 or more CAG repeats. While there are inconsistencies in defining at which point polyglutamine *ATXN2* expansions increase ALS risk in previously published studies, these discrepancies may be due to the diverse ethnic backgrounds investigated in existing research and to unrecognized population substructures in cases and controls.

Sizing of CAG repeats can be challenging. Because none of the studies reported a common sizing standard, we assessed the quality of sizing by examining the frequency distribution of alleles. All studies showed the expected distribution with 22 repeats as the most common allele, followed by alleles with 23 and 27 repeats.^{1,21,22} The frequency of the 27-repeat allele in individual studies, however, showed significant variability ranging from 0.2%¹³ to 3.6%.¹² Similar observations hold true for the 28- and 29-repeat alleles. The observed variation in frequency likely represents unrecognized population substructure and can, in retrospect, explain the spurious association of increased ALS risk for these alleles. It is also possible that these alleles have an increased risk in specific populations only.

Although the stated objectives of the studies included in our meta-analysis were similar, there were some differences in the definition and composition of ALS and control populations. The ALS and control cohorts were recruited from different geographical and ethnic regions including the United States,⁶ French Canada/Canada,¹⁰ Belgium and the Netherlands,¹² Germany,¹¹ Italy,^{9,16} mainland China,¹⁵ Turkey,¹³ and Flanders-Belgium.¹⁴ Additionally, the source and type of

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Figure 2. Odds Ratios (ORs) for Individual Ataxin 2 Gene (*ATXN2*) Alleles With 30 to 33 CAG Repeats Compared With Alleles With 23 or Fewer CAG Repeats

A 30 CAG	peats relative to the baseline (≤23 repeats)
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Source	OR (95% CI)
Elden et al, ⁶ 2010	9.81 (0.53-182.24)
Corrado et al, ⁹ 2011	0.40 (0.02-9.94)
Daoud et al, ¹⁰ 2011	2.59 (0.27-24.93)
Gispert et al, ¹¹ 2012	7.33 (0.76-70.56)
Van Damme et al, ¹² 2011	1.80 (0.65-4.95)
Gellera et al, ¹⁶ 2012	2.52 (0.26-24.28)
Van Langenhove et al, ¹⁴ 2012	23.60 (2.13-261.94)
Liu et al, ¹⁵ 2013	0.20 (0.05-0.79)
Lahut et al, ¹³ 2012	(Excluded)
Overall (1 ² =61.1%, P=.01)	1.55 (0.88-2.73)



B 31 CAG repeats relative to the baseline (≤23 repeats)

Source	OR (95% CI)
Elden et al, ⁶ 2010	3.81 (0.79-18.38)
Corrado et al, ⁹ 2011	8.48 (0.44-164.57)
Daoud et al, ¹⁰ 2011	4.31 (0.50-36.99)
Gispert et al, ¹¹ 2012	0.81 (0.03-20.01)
Van Damme et al, ¹² 2011	1.08 (0.40-2.88)
Gellera et al, ¹⁶ 2012	5.88 (0.72-47.90)
Lahut et al, ¹³ 2012	8.19 (0.33-201.70)
Van Langenhove et al, ¹⁴ 2012	3.92 (0.16-96.69)
Liu et al, ¹⁵ 2013	6.19 (0.35-109.90)
Overall (1 ² =0.0%, P=.62)	2.70 (1.47-4.93)



1.0

OR (95% CI)

Weight, % 11.22 10.60 25.15 6.82 11.29 6.28 12.73 15.90 0.00 100.00

870

C 32 CAG repeats relative to the baseline (≤23 repeats)

Source	OR (95% CI)
Elden et al, ⁶ 2010	18.52 (1.07-321.11)
Corrado et al, ⁹ 2011	8.48 (0.44-164.57)
Daoud et al, ¹⁰ 2011	5.18 (0.62-43.08)
Gispert et al, ¹¹ 2012	17.10 (0.88-331.35)
Van Damme et al, ¹² 2011	22.66 (1.33-386.78)
Lahut et al, ¹³ 2012	13.66 (0.65-285.29)
Gellera et al, ¹⁶ 2012	10.93 (0.61-194.22)
Liu et al, ¹⁵ 2013	5.23 (0.29-94.74)
Van Langenhove et al, ¹⁴ 2012	(Excluded)
Overall (1 ² =0.0%, P=.99)	11.09 (4.16-29.57)

D 33 CAG repeats relative to the baseline (≤23 repeats)



0.00115

Data are shown for 30 (A), 31 (B), 32 (C), and 33 (D) CAG repeats relative to the baseline of 23 or fewer CAG repeats. Horizontal lines indicate 95% CI; arrow (D) indicates an off-scale 95% CI. Dashed line indicates the pooled OR, with the width of the diamond representing the 95% CI. Square size is representative of the size or weight of the individual study.

controls differed from one study to the next; controls were defined as neurologically normal,^{6,11,12} neurologically normal

without any known history of neurological disorders,^{13,15} neurologically normal matched for age,¹⁰ neurologically normal

matched for age and ethnicity, ^{9,10,15} and neurologically normal with exclusion of incipient Parkinson disease through midbrain sonography.¹¹

As shown in both Figure 1 and the eTable in the Supplement, several studies identified pathogenic SCA2 alleles with 34 or more repeats in patients with ALS. In total, 15 patients with ALS and full-length pathological SCA2 repeats (\geq 34) presented with a motor neuron syndrome as opposed to a cerebellar syndrome. A recent study described a family with coexisting SCA2 and ALS.²⁴ The proband with *ATXN2* CAG repeats of 40 and 22 presented with degenerative ataxia, while her uncle had an upper and lower motor neuron syndrome with CAG repeats of 39 and 22. Familial ALS, caused by mutant *ATXN2* alleles, was also observed in a Cuban population.²⁵ This study, in addition to our findings, further supports the notion that expanded *ATXN2* CAG repeats can give rise to several neurological disorders with a wide range of phenotypes.

It is not known what causes the shift from a typical cerebellar phenotype to a rare motor phenotype in SCA2. In principle, genetic, environmental, and stochastic factors could play a role. The occurrence of individuals with the typical cerebellar phenotype and the ALS phenotype in the same pedigree argues against importance of cis-acting factors. Nonallelic modifiers (genetic background) cannot be excluded, but the presence of the SCA2 ALS phenotype in different ethnic groups may make this mechanism less likely. The relative rarity of the ALS phenotype suggests to us that stochastic events such as expansion of long-normal ATXN2 alleles in motor neurons during mitosis or DNA repair, both of which have genetic, epigenetic, or environmental underpinnings, may explain the exclusive motor neuron phenotype in some individuals. The possibility that only ataxin 2 molecules with a specific and narrow window of polyglutamine repeat lengths have an abnormal interaction with TDP-43 seems less likely.⁶ At this point, however, experimental data are lacking both in model systems and in human patients.

The *ATXN2* alleles with 32 and 33 repeats are considered extremely rare and have been associated with very lateonset cerebellar ataxia for 32 repeats in the homozygous state in one publication²⁶ (and by personal observation by one of us [S.M.P.]) and 33 repeats in the heterozygous state in another.² Consistent with that notion, only 1 allele with 32 repeats and no alleles with 33 repeats were found among approximately 15 000 control alleles. Among approximately 12 000 ALS alleles, there were 43 alleles with 32 repeats and 15 alleles with 33 repeats.

In ALS, it is highly probable that both known and unknown susceptibility genes interact with environmental exposures to modulate disease risk.²⁷ In addition to longnormal *ATXN2* repeats, several other genes and environmental factors have been shown to have an association with ALS. Careers such as farming (OR = 2.0; 95% CI, 1.1-3.5)²⁸ and some electrical occupations (OR = 2.3; 95% CI, 1.29-4.09)²⁹ may be related to an increased risk of ALS.

Additionally, having ever smoked a cigarette (OR = 2.0; 95% CI, 1.3-3.2)³⁰ or having a high intake of dietary fat (OR = 2.78; 95% CI, 0.9-8.0)³¹ have shown association with ALS; however, dietary fiber (OR = 0.3; 95% CI, 0.1-0.7)³¹ may have a preventive effect. Thus, ORs of 2.70, 11.09, and 5.76 for 31, 32, and 33 *ATXN2* repeats, respectively, represent significant risk factors for ALS.

Conclusions

This meta-analysis provides a new framework for interpretation and counseling of patients with regard to ATXN2 alleles. Alleles with up to 30 CAG repeats can be considered normal with no apparent increased risk for neurodegenerative disease. In fact, alleles of 27 and 28 repeats had a slight but significantly reduced risk of ALS. Our analysis confirms that there is an increased risk for ALS with alleles of 31, 32, and 33 repeats. It is not known whether repeat interruptions modify this risk. Counseling also needs to include information that longnormal alleles show increased meiotic instability and can expand to full-mutation alleles in offspring. For alleles of 34 or more repeats, most individuals will develop a cerebellar degeneration but some may present with motor neuron disease (Figure 1). Of note, many patients with SCA2 presenting with cerebellar dysfunction will develop symptoms and signs of motor neuron disease late in the course of their illness.

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Study concept and design: Pulst.

Acquisition, analysis, or interpretation of data: All authors. Drafting of the manuscript: Neuenschwander, Thai,

Pulst.

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REFERENCES

1. Pulst SM, Nechiporuk A, Nechiporuk T, et al. Moderate expansion of a normally biallelic trinucleotide repeat in spinocerebellar ataxia type 2. *Nat Genet*. 1996;14(3):269-276. 2. Fernandez M, McClain ME, Martinez RA, et al. Late-onset SCA2: 33 CAG repeats are sufficient to cause disease. *Neurology*. 2000;55(4):569-572.

 Pulst SM, Santos N, Wang D, et al.
Spinocerebellar ataxia type 2: polyQ repeat variation in the CACNA1A calcium channel modifies age of onset. *Brain.* 2005;128(pt 10):2297-2303.

4. Scherzed W, Brunt ER, Heinsen H, et al. Pathoanatomy of cerebellar degeneration in spinocerebellar ataxia type 2 (SCA2) and type 3 (SCA3). *Cerebellum*. 2012;11(3):749-760.

5. Lastres-Becker I, Rüb U, Auburger G. Spinocerebellar ataxia 2 (SCA2). *Cerebellum*. 2008; 7(2):115-124.

 Elden AC, Kim HJ, Hart MP, et al. Ataxin-2 intermediate-length polyglutamine expansions are associated with increased risk for ALS. *Nature*. 2010;466(7310):1069-1075.

7. Lagier-Tourenne C, Cleveland DW. Rethinking ALS: the FUS about TDP-43. *Cell*. 2009;136(6): 1001-1004.

8. Pesiridis GS, Lee VM, Trojanowski JQ. Mutations in TDP-43 link glycine-rich domain functions to amyotrophic lateral sclerosis. *Hum Mol Genet*. 2009;18(R2):R156-R162.

9. Corrado L, Mazzini L, Oggioni GD, et al. ATXN-2 CAG repeat expansions are interrupted in ALS patients. *Hum Genet*. 2011;130(4):575-580.

 Daoud H, Belzil V, Martins S, et al. Association of long ATXN2 CAG repeat sizes with increased risk of amyotrophic lateral sclerosis. *Arch Neurol.* 2011; 68(6):739-742.

11. Gispert S, Kurz A, Waibel S, et al. The modulation of amyotrophic lateral sclerosis risk by ataxin-2 intermediate polyglutamine expansions is a specific effect. *Neurobiol Dis.* 2012;45(1):356-361.

12. Van Damme P, Veldink JH, van Blitterswijk M, et al. Expanded ATXN2 CAG repeat size in ALS identifies genetic overlap between ALS and SCA2. *Neurology*. 2011;76(24):2066-2072.

13. Lahut S, Ömür Ö, Uyan Ö, et al. ATXN2 and its neighbouring gene SH2B3 are associated with increased ALS risk in the Turkish population. *PLoS One*. 2012;7(8):e42956.

14. Van Langenhove T, van der Zee J, Engelborghs S, et al. Ataxin-2 polyQ expansions in FTLD-ALS spectrum disorders in Flanders-Belgian cohorts. *Neurobiol Aging*. 2012;33(5):e17-e20.

15. Liu X, Lu M, Tang L, Zhang N, Chui D, Fan D. ATXN2 CAG repeat expansions increase the risk for Chinese patients with amyotrophic lateral sclerosis. *Neurobiol Aging*. 2013;34(9):e5-e8.

16. Gellera C, Ticozzi N, Pensato V, et al. *ATAXIN2* CAG-repeat length in Italian patients with amyotrophic lateral sclerosis: risk factor or variant phenotype? implication for genetic testing and counseling. *Neurobiol Aging*. 2012;33(8):e15-e21.

17. Lane PW. Meta-analysis of incidence of rare events. *Stat Methods Med Res.* 2013;22(2):117-132.

18. Friedrich JO, Adhikari NK, Beyene J. Inclusion of zero total event trials in meta-analyses maintains analytic consistency and incorporates all available data. *BMC Med Res Methodol*. 2007;7:5.

19. Mark A, Kaelin MB. Attributable risk applications in epidemiology. http://www .collegeboard.com/prod_downloads/yes/4297 _MODULE_17.pdf. Accessed March 28,2014.

20. Brooks BR, Miller RG, Swash M, Munsat TL; World Federation of Neurology Research Group on Motor Neuron Diseases. El Escorial revisited: revised criteria for the diagnosis of amyotrophic lateral sclerosis. *Amyotroph Lateral Scler Other Motor Neuron Disord*. 2000;1(5):293-299.

21. Sanpei K, Takano H, Igarashi S, et al. Identification of the spinocerebellar ataxia type 2 gene using a direct identification of repeat expansion and cloning technique, DIRECT. *Nat Genet*. 1996;14(3):277-284.

22. Imbert G, Saudou F, Yvert G, et al. Cloning of the gene for spinocerebellar ataxia 2 reveals a locus with high sensitivity to expanded CAG/glutamine repeats. *Nat Genet*. 1996;14(3):285-291.

23. Figueroa KP, Farooqi S, Harrup K, Frank J, O'Rahilly S, Pulst SM. Genetic variance in the spinocerebellar ataxia type 2 (*ATXN2*) gene in children with severe early onset obesity. *PLoS One*. 2009;4(12):e8280.

24. Tazen S, Figueroa K, Kwan JY, et al. Amyotrophic lateral sclerosis and spinocerebellar ataxia type 2 in a family with full CAG repeat expansions of *ATXN2*. *JAMA Neurol*. 2013;70(10): 1302-1304.

25. Laffita-Mesa JM, Rodríguez Pupo JM, Moreno Sera R, et al. De novo mutations in ataxin-2 gene and ALS risk. *PLoS One.* 2013;8(8):e70560.

26. Almaguer-Mederos LE, Falcón NS, Almira YR, et al. Estimation of the age at onset in spinocerebellar ataxia type 2 Cuban patients by survival analysis. *Clin Genet*. 2010;78(2):169-174.

27. Al-Chalabi A, Hardiman O. The epidemiology of ALS: a conspiracy of genes, environment and time. *Nat Rev Neurol.* 2013;9(11):617-628.

28. McGuire V, Longstreth WT Jr, Nelson LM, et al. Occupational exposures and amyotrophic lateral sclerosis: a population-based case-control study. *Am J Epidemiol.* 1997;145(12):1076-1088.

29. Noonan CW, Reif JS, Yost M, Touchstone J. Occupational exposure to magnetic fields in case-referent studies of neurodegenerative diseases. *Scand J Work Environ Health*. 2002;28(1): 42-48.

30. Nelson LM, McGuire V, Longstreth WT Jr, Matkin C. Population-based case-control study of amyotrophic lateral sclerosis in western Washington State, I: cigarette smoking and alcohol consumption. *Am J Epidemiol*. 2000;151(2):156-163.

31. Nelson LM, Matkin C, Longstreth WT Jr, McGuire V. Population-based case-control study of amyotrophic lateral sclerosis in western Washington State, II: diet. *Am J Epidemiol*. 2000;151 (2):164-173.